

Defining the New Normal in Quantitative Western Blot Data

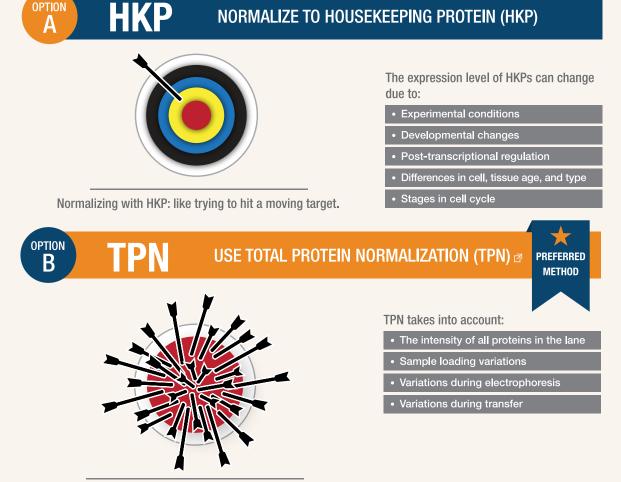
By Geetha Yadav and Kenneth Oh

Do you know that journals like the JBC strongly caution against the use of housekeeping proteins for normalization? Most journals now demand rigorous standards for presentation of western blotting data. While housekeeping proteins (HKP) have traditionally been used for decades, the problems with their usage have come to the fore only recently. More validation is required for presenting data using HKP. Normalization using total proteins present in the sample has gained significant traction and has proved to be a reliable method for normalization of western blots. Here we illustrate why the **total protein normalization (TPN)** method is superior and the way to go for western blotting.



DEFINING THE NEW NORMAL IN QUANTITATIVE WESTERN BLOT DATA

HOW DO I ACCURATELY QUANTITATE THE CHANGES IN MY PROTEINS?



Normalizing with total protein: multiple arrows covering the entire target area.

TPN PROVIDES ACCURATE QUANTITATION OF TARGET PROTEINS.



Total protein normalization is the most reliable method of reporting quantitative western blot data.

SEVERAL GROUPS HAVE EXPRESSED **CONCERNS REGARDING WESTERN BLOT DATA NORMALIZATION:**



TPN is the preferred method of normalization for:

- Stem cell research
- Neuroscience research
- · Alzheimer's research
- Developmental biology
- Aging studies



Science

NIH workshops and meetings



JOURNALS LIKE JOURNAL OF BIOLOGICAL CHEMISTRY PREFER TPN FOR SUBMITTING WESTERN BLOT DATA FOR PUBLICATION.

JBC editors recommend TPN for normalizing western blots.

It is typically better to normalize western blots using total protein loading as the denominator.

We prefer that signal intensities are normalized to total protein by staining membranes with Coomassie Blue, Ponceau S, or other protein stains, and we strongly caution against the use of housekeeping proteins for normalization, unless there is clear demonstration that expression of the housekeeping protein is unaffected by experimental treatments.

Fosang AJ and Colbran RJ (2015). Transparency is the key to quality. J Biol Chem 290, 29,692-29,694.



- sunoblots (Western blotti cy on image manipulation

dies reporting semi-quantitative analyses of immunoblots, authors should clearly explain how p For studies reporting semi-quartitative analyses or immunotoion, autono should charry explain how quant data were obtained, whether signal intensity has a linear relationship with antigen loading, and how protein was normalized among lanes. Note that some detection methods including detection of enhanced chemiluminescence using X-ray film have a very limited linear range. Normalization of signal intensity to to protein loading (assessed by staining membranes using Coomassie blue, Ponceau S or other protein stains) end obtaining passessed up samming instroadents using Colonausie using, revealed a to using the end. "Nouse-teaping" proteins should not be used for normalization without evidence that ipulations do not affect their expression. Signals obtained using antibodies specific for pho aid be normalized to the total protein level of the target protein. nce that exp

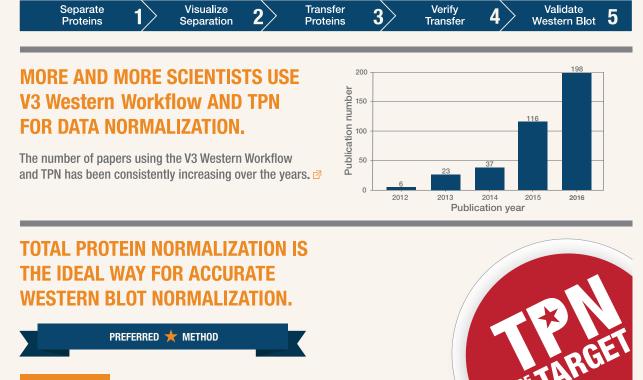
http://www.jbc.org/site/misc/ifora.xhtml

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BIO-RAD'S V3 Western Workflow [™] INCORPORATES TPN FOR NORMALIZATION OF WESTERN BLOT DATA. • Qua lane • Norr prote

V3 Western Workflow includes:

- Stain-free visualization of total proteins on gel after electrophoresis
- Quantitation of total proteins in a lane after transfer
- Normalization of bands to total proteins by specialized software



learn more

about TPN and Bio-Rad's V3 Western Workflow.

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